

PROSITE and HAMAP

Remember to carefully read the documentation available on the web pages, as a lot of useful information can be found there.

1. The PROSITE database and ScanProsite

The aim of this exercise is to explore and understand the PROSITE database.

General information in a PROSITE entry

First have a look at the following PROSITE entry: [PS50235](#).

- Is the descriptor PS50235 a PROSITE pattern or a profile? How do you distinguish one from the other?
- Assuming that this descriptor matched your sequence, would you necessarily believe the result? And if it did not match?
- What is the predicted function of proteins matching PS50235? (*Hint* - check the PDOC documentation)
- Is PS50235 related to other PROSITE entries? If yes, are they patterns or profiles? What is the quality of these signatures? (*Hint* - check the PDOC documentation)

Sequence analysis using PROSITE

Analyze the following sequence using [ScanProsite](#):

```
>seq1
MELRVLLCWASLAAALEETLLNNTKLETADLKWVTFPQVDGQWEELSGLDDEEQHSVRTYEV
CDVQRAPGQAHWLRGTGWVPRRGAVHVYATLRFTMLECLSLPRAGRCKETFTVFYYESDA
DTATALTPAWMENPYIKVDTVAAEHLTRKRPGAEATGKVNKTLRLGPLSKAGFYLAQD
QGACMALLSLHLFYKKAQLTVNLTRFPETVPRELVVPVAGSCVVDVAVPAPGSPSPSLYCR
EDGQWAEQPVTCGSCAPGFEEAAGNTKCRACAQGTFFKPLSGEGSCQPCPANSHTIGSA
VCQCRVGYFRARTDPRGAPCTTPPSAPRSVVSRLNGSSLHLEWSAPLES GGREDLTYALR
CRECRPGGSCAPCGGDLTFDPGPRDLVEPWVVVRGLRPDFTYTFEVTALNGVSSLATGPV
PFEPVNVTTDREVPPAVSDIRVTRSSPSSLSLAWAVPRAPSGAVLDYEVKYHEKGAEGPS
SVRFLKTSENRAELRGLKRGASYLVQVRARSEAGYGFPGQEHHSQTQLDESEGWREQLAL
IAGTAVGVVLLVVLVIVVAVLCLRKQSNGREAEYSKDHGQYLIGHGTKVYIDPFTYEDPN
EAVREFAKEIDVSYVKIEEVI GAGEFGEVCRGRLKAPGKKECVAIKTLKGGYTERQRRE
FLSEASIMGQFEHPNIIRLEGVVTSNMPVMILTEFMENGALDSFLRLNDGQFTVIQLVGM
LRGIASGMRYLAEMSYVHRDLAARNILVNSNLVCKVSDFGLSRFLLEENSSDPTYTSSLGG
```

KIPIRWTAPEAIAFRKFTSASDAWSYGIVMWEVMSFGERPYWMSNQDVINAIEQDYRLP
PPDCPTSLHQLMLDCWQKDRNARPRFPQVVSALDKMIRNPASLKIVARENGGASHPLLD
QRQPHYSAFGSVGEWLRAIKMGRYEESFAAAGFGSFELVSQISAEDLLRIGVTLAGHQKK
ILASVQHMKSQAKPGTGGTGGPAPQY

- What is the domain composition of the protein? What is its function?
- Can you identify potential binding sites using PROSITE? (*Hint* - Move the mouse on the sequences and images in the results page to highlight information)

Now look at this sequence from a patient with a cardio-vascular disease.

>seq2

MELRVLLCWASLAAALEETLLNFKLETADLKWVTFPQVDGQWEELSGLDDEEQHSVRTYEV
CDVQRAPGQAHLWLRGTGWVPRRGAVHVVYATLRFMTLECLSLPRAGRSCKETFTVFYYESDA
DTATALTPAWMENPYIKVDTVAAEHLTRKRPGAEATGKVVNKTLLRLGPLSKAGFYLAQD
QGACMALLSLHLFYKKCAQLTVNLTRFPETVPRELVVPVAGSCVVDVAVPAPGSPSPSLYCR
EDGQWAEQPVTCGSCAPGFEAAEGNTKCRACAQGTFKPLSGEGSCQPCPANSHTIGSA
VCQCRVGYFRARTDPRGAPCTTPPSAPRSVVSRLNGSSLHLEWSAPLES GGREDLTYALR
CRECRPGGSCAPCGDLTFDPPGPRDLVEPWVVVRGLRPFDTYTFEVTALNGVSSLATGPV
PFEPVNVTTDREVPPAVSDIRVTRSSPSSLSLAWAVPRAPSGAVLDYEVKYHEKGAEGPS
SVRFLKTSENRAELRGLKRGASYLVQVRARSEAGYGPFGQEHHSQTQLDESEGWREQLAL
IAGTAVGVVLLVVLVIVVAVLCLRKQSNGREAEYSKDHGQYLIHGHTKVYIDPFTYEDPN
EAVREFAKEIDVSYVKIEEVI GAGEFGEVCRGLKAPGKKECVAI STLKGGYTERQRRE
FLSEASIMQFEHPNII RLEGVVTNSMPVMILTEFMENGALDSFLRLNDGQFTVIQLVGM
LRGIASGMRYLAEMSIVHRDLAARNILVNSNLVCKVSDFGLSRFLEENSSDPTYTSSLGG
KIPIRWTAPEAIAFRKFTSASDAWSYGIVMWEVMSFGERPYWMSNQDVINAIEQDYRLP
PPDCPTSLHQLMLDCWQKDRNARPRFPQVVSALDKMIRNPASLKIVARENGGASHPLLD
QRQPHYSAFGSVGEWLRAIKMGRYEESFAAAGFGSFELVSQISAEDLLRIGVTLAGHQKK
ILASVQHMKSQAKPGTGGTGGPAPQY

- What is the difference between this sequence and the previous one?
- Can you suggest a possible underlying mechanism for this patient's disease?
- Complement your answer using information you can retrieve using [UniProt](#).

Hint - a [literature](#) reference.

2. The HAMAP database and HAMAP-Scan

The HAMAP-Scan "Scan" mode can be used to classify protein sequences using HAMAP profiles, while the HAMAP-Scan "Scan & Annotate" mode also provides annotation covering individual sequences and complete proteomes. Both are available [here](#).

To save time we have annotated a number of proteomes and individual sequences for you. The corresponding results from each of these sequences can be retrieved from the [HAMAP-Scan results page](#) using the access codes provided in the following tables.

2a. HAMAP classification and (conditional) annotation

Individual sequences:

Species name	Taxonomic identifier	HAMAP-Scan access code
<i>Escherichia coli</i> (strain K12)	83333	FGV
<i>Bacillus cereus</i> var. <i>anthracis</i> (strain CI)	637380	IIO

Retrieve the annotations for the individual sequences of *Escherichia coli* (strain K12) and *Bacillus cereus* var. *anthracis* (strain CI) shown below at the [HAMAP-Scan results page](#). At the same time, copy/paste each of these sequences into the HAMAP-Scan search box [here](#) and use the simple "Scan" mode to search all HAMAP profiles in real time for matches to each of the sequences.

>*Escherichia coli*

```
MLKIFNTLTRQKEEFKPIHAGEVGMVCGITVYDLCHIGHGRTFVAFDVVARYLRFLGYK
LKYVRNITDIDDKI I KRANENGESFVAMVDRMIAEMHKDFDALNILRPDMEPRATHHIAE
I IELTEQLIAKGHAYVADNGDVMFDVPTDPTYGVLSRQDLQAGARVDVDDKRNPM
FVLWKMSKEGEPSPWSPWGAGRPGWHIECSAMNCKQLGNHFDIHGGGSDLMFPHHENEIA
QSTCAHDGQYVNYWMHSGMVMVDREKMSKSLGNFFTVRDVLKYYDAETVRYFLMSGHYRS
QLNYSEENLKQARAALERLYTALRGTDKTVAPAGGEAFEARFIEAMDDDFNTPEAYSVLF
DMAREVNRLKAEDMAAANAMASHLRKLSAVLGLLEQEPEAFLOSGAQADDSEVAEIEALI
QQRLDARKAKDWAAADAARDRLNEMGIVLEDGPQGTWRRK
```

>*Bacillus cereus* var. *anthracis*

```
MTIHIYNTLTRQKEEFTPLEENKVKMYVAGPTVYNYI HIGNARPPMVFDTVRRYLEYKGY
DVQYVSNFTDVDDKLI KAANELGEDVPTIADRFVEAYFEDVTALGCKHATVHPRVTENMD
I IIEFIQELVNKGYAYESEGDVYFRTKEFEGYKLSHQPIADLRHGARIEVGEKKQDPLD
FALWKAKEGEI FWESPWGQGRPGWHIECSAMARKYLGDTIDIHAGGQDLAFPHHENEIA
QSEALTGKTFARYWMHNGYININNEKMSKSLGNFILVHDI IKQYDPQLIRFFMFSVHYRH
PINFSEELLQSTNGLERIKTAYGNLKHRESSTDLTDHNEKWLADLEKFQTAFFEEAMND
DFNTANAITELYNVANHANQYLLEEHTSTVVIEAYVKQLETFLFDILGLELAQEELLDEEI
EELIQKRIEARKNRDFALSQIRDDLKDRNI ILEDTAQGTWRWKR
```

- Compare the output of the HAMAP-Scan "Scan" and "Scan & Annotate" modes. What information is specific to each?
- Now compare the annotation of the two sequences from the HAMAP-Scan "Scan & Annotate". What differences can you see in the resulting annotation?
- Can you deduce the reasons for these differences? Hint: examine the 'cases' in the HAMAP rule used to produce these annotation (the rule can be identified from the "DR HAMAP" line) as well as the annotation "warnings" provided for each sequence (in the "INTERNAL SECTION"). Can you see why it is necessary to specify a taxonomic node when performing "Scan & Annotate"?

2b. HAMAP proteome annotation

At the end of 2019, a novel coronavirus (nCoV) of animal origin started infecting humans, initiating a severe outbreak in China. On January 30th, 2019-nCoV was designated a global health emergency by the WHO. On February 11th, the WHO called the disease caused by the virus COVID-19, and the virus itself was named Severe Acute Respiratory Syndrome-related coronavirus 2 or SARS-CoV-2 by the International Committee on Taxonomy of Viruses (ICTV).

SARS-CoV-2 belongs to the large family of Coronaviridae. This genus comprises mainly vertebrate respiratory viruses, including HCoV-OC43, which is responsible for 10% of common colds, and SARS, which caused an epidemic in 2003. The novel coronavirus genome has been sequenced.

SARS-CoV-2 protein sequences from the current public health emergency have been made available as a [pre-release dataset](#) on the UniProt FTP site.

Download the 14 protein sequences of SARS-CoV-2 in fasta from UniProt [here](#) and submit them to [HAMAP-Scan](#)

- What proportion of the proteome has been annotated by HAMAP?
- Do the results tell you more about the taxonomy of the strain?

Next, retrieve the annotations for the SARS-CoV-2 sequences from the [HAMAP-Scan results page](#) using the access codes provided below.

SARS-CoV-2:

Species name	Taxonomic identifier	HAMAP-Scan access code
<i>Coronaviridae</i>	11118	FGU

- What proportion of the proteome has been annotated by HAMAP?
- How many rules for coronavirus proteins in general and for betacoronavirus proteins do we have in HAMAP? What do you think are the reasons that not more proteins were annotated?
- Can you explain why some proteins were not annotated, although there seems to be a HAMAP-rule available? And how could we improve the situation? (*Hint* Check the "Scan"-Results you did before and look for the SPIKE protein (MF_04099))

For advanced students

3. Build your own pattern

You are working with a family of proto-oncogene proteins and have identified a potential functional region that is conserved in several related proteins. You have built a multiple sequence alignment of this region from these proteins, and now you would like to identify other proteins having this signature.

- Build a Pattern from the MSA using the PROSITE syntax (you can find it [here](#)). *Hint* - you can use [weblogo](#) to guide you.

```
Seq1 WFFKGIADKDAERHLLA
Seq2 WFFKNLEQKDAEARLLA
Seq3 WFFKR---KDAERQLLA
Seq4 WFFGTI---DAERQLLA
Seq5 WFFKDIPTKDAERQLLA
Seq6 WYFG----RESERLLLA
Seq7 WYFGKIPLKDAERQLLA
Seq8 WYFGKLRKDAERLLLL
```

The first thing to do now is to check the quality of your pattern.

- How will you do that? *Hint* - look at the [ScanProsite](#) page.

Search the UniProtKB/Swiss-Prot database with your pattern using [ScanProsite](#).

- What can you say about the proteins matching your pattern?

Repeat the exercise with the following sequences:

```
seq1 ERGLAAAR
seq2 DRVSCLIR
seq3 DRLGSGGR
seq4 ERAALILR
seq5 ERIVVTVR
```

- How easy is it to build a good quality pattern? What is the source of any difficulties?

4. MyHits Tutorial

More practicals on how to use [MyHits](#) (a SIB resource where you can build your own profiles and HMMs):

- Go to the page <http://myhits.vital-it.ch/doc/tutorial-psi.html> if you want to do some more practicals on PSI-BLAST.

- Go to this page <http://myhits.vital-it.ch/doc/tutorial-domain.html> for more practicals in domain hunting.